

A Joint Usage/Research Center approved
by the Japanese Ministry of Education,
Culture, Sports, Science and Technology
and one of the WOAHP Collaborating Centres

National Research Center for Protozoan Diseases



National Research Center for Protozoan Diseases
Obihiro University of Agriculture and Veterinary Medicine



2026

Obihiro University of
Agriculture and Veterinary Medicine

Mission

Our mission, as the only MEXT Joint Usage/Research Center of Protozoan Diseases in Japan, is to promote cutting-edge research on the control of protozoan diseases in humans and livestock. This research is conducted to contribute to the health and welfare of human beings as well as to the global issue of food safety and security. This will be accomplished through cooperation with domestic and foreign universities, relevant ministries and international institutes.



Message from the Director General

Noboru Inoue

The National Research Center for Protozoan Diseases (NRCPD) of Obihiro University of Agriculture and Veterinary Medicine was established as an inter-university joint-use facility in 1990, certified as a national joint-use facility in 2000, and then as a joint-use facility/research center in 2009 by the Ministry of Education, Culture, Sports, Science and Technology. In addition, in terms of collaborative activities with related international organizations, our two professors were appointed as Reference Laboratory (RL) experts by the World Organization for Animal Health (WOAH) in 2007, and the following year, NRCPD was recognized as the first WOAH Collaborating Center in the world in the field of protozoan diseases. In 2017, the WOAH RL developed an international standard diagnostic testing system by obtaining ISO/IEC 17025 accreditation for the diagnostic tests provided by the WOAH RL. In collaboration with WOAH and other relevant international organizations, we will continue to contribute to the global surveillance and control of protozoan diseases through academic research and international human resource development. As an example, since 1995 for 30 years, with the support of the Japan International Cooperation Agency (JICA), NRCPD has invited experts from emerging and developing countries who are involved in zoonotic

disease control, mainly protozoan diseases, and has conducted group training courses. Nearly 233 trainees have so far completed the course and are now playing a central role in veterinary and livestock administration and infectious disease research and education in their respective countries. Our network of international researchers consisting of these graduates has become an important human resource in the internationalization of education and research activities at the University as well as at NRCPD.

The damage caused to humans and animals by protozoan diseases is immeasurable. Protozoan parasites are tough enemies with a high similarity to host cells and sophisticated survival strategies such as antigenic variations, immune disruption, and complex life cycles. Unfortunately, there are still few prophylactic vaccines or safe treatments available. Therefore, the development of inexpensive diagnostic, prophylactic, and therapeutic methods that can be practically applied in livestock farms around the world is an urgent issue on a global scale. NRCPD will contribute to One Health issue resolution and promotion by actively promoting international collaborative research based on our global network of protozoan disease researchers.

International Contribution/Development of Human Resources

The National Research Center for Protozoan Diseases, in collaboration with the Japan International Cooperation Agency (JICA), has provided training programs on zoonotic protozoan diseases to backbone and senior administrative technical researchers from developing countries, especially Asian and African nations, since 1995. Over 233 graduates working as advanced technical researchers perform zoonoses measures in their respective countries and act as important counterparts for the international joint research project. We also provide graduates the opportunity to join our research center for 3-6 months to re-train their professional skills and develop collaborative research projects with us. We accept 3~5 re-trainees every fiscal year.



Providing technical assistance for the diagnosis of protozoan diseases in Vietnam



Opening ceremony for a JICA advanced training course



Workshop for JICA trainee



Providing technical assistance for the diagnosis of malaria in the Philippines

International Joint Research Center

Many protozoan diseases persistently infect livestock, causing anemia and miscarriages and worsening their chronic health condition. However, accurate means of diagnosis, prophylactic vaccines and safe specific medicines are not available for protozoan diseases. The detrimental effects of protozoan diseases on livestock must therefore be resolved on a global scale as quickly as possible. The National Research Center for Protozoan Diseases, as the international joint research center for protozoan diseases of animals in the world, has clarified the distribution and damage caused by these diseases by conducting a large-scale epidemiological survey using diagnostic technology developed in-house. Moreover, we train

young specialists throughout the world through these kinds of activities at the international joint research center. The center also continues to develop and make contributions to research on protozoan diseases, as well as maintaining an early recognition system and thereby contributing to the prevention of these diseases in livestock and thus to the development of the international livestock industry.

Cooperation countries:

The United States of America, Mongolia, China, Taiwan, South Korea, Philippines, Thailand, Vietnam, Indonesia, Sri Lanka, Egypt, Kenya, Uganda, Tanzania, Benin, Burkina Faso, South Africa, Turkey, India, Germany, Poland, Argentina, Mexico, Paraguay and Kyrgyzstan



Investigation for tsetse fly in Zambia



Epidemiological survey for livestock protozoan diseases in Vietnam



Epidemiological survey for livestock protozoan diseases in Mongolia



Investigation of ticks that transmit protozoa in China

WOAH Reference Laboratory and Collaborating Centre

The World Organization for Animal Health (WOAH), an international organization with 183 member countries, aims to ensure the security and safety of livestock and livestock products. They certify the world's cutting-edge research institutes and specialists as collaborating centres (CC) or reference laboratories (RL). These institutes then utilize the results of the latest research for the development of new diagnostic methods for infectious diseases in animals and are responsible for the international standardization of vaccines. In recent years, WOAH has also emphasized the contribution of CCs to developing countries so that diagnostic

technology for infectious diseases and livestock sanitation standards can be improved in these countries. In June 2007, our two professors were designated as WOAH RL experts (bovine babesiosis, equine piroplasmosis and surra). In May 2008, the National Research Center for Protozoan Diseases (NRCPD) was designated as the first WOAH CC in Asia. NRCPD is highly expected to contribute for the control of zoonotic protozoan diseases in the world. Moreover, "protozoan DNA testing of bovine babesiosis, equine piroplasmosis and surra by PCR methods in accordance with WOAH Manual" provided by WOAH RLs has been certified as ISO/IEC 17025:2017



Non-tsetse transmitted animal trypanosomoses (NTTAT) specialist conference at the WOAH headquarters



Joint hosting of the WOAH Regional Workshop



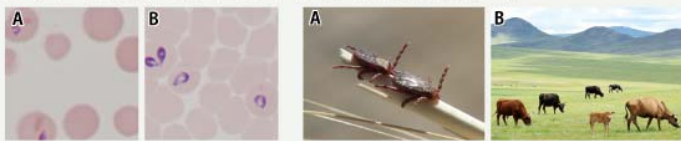
Plate declaring the National Research Center for Protozoan Diseases an WOAH Collaborating Centre



**Professor
Naoaki Yokoyama**

Bovine and equine piroplasmosis (babesiosis and theileriosis) caused by species of genera *Babesia* and *Theileria* is characterized by fever and anemia. The disease usually results in severe economic losses in cattle and horse industry worldwide. Therefore, control and

prevention of bovine and equine piroplasmosis through surveillance and risk factor identification are vital. As WOAHP reference laboratories for bovine babesiosis and equine piroplasmosis, we survey the cattle and horses on a global scale to ascertain the current epidemiological status of *Babesia* and *Theileria* species, determine their genetic diversity, identify the risk factors, and discover novel parasite species with clinical importance. We also develop methods and scientific tools to aid better research and diagnosis. Moreover, we provide diagnostic services and materials to WOAHP member countries to improve control and preventive efforts, particularly to prevent the spread of infections through the transnational movement of animals. To accelerate our goal of minimizing bovine and equine piroplasmosis, we maintain a robust international collaborative network and welcome postgraduate students and young postdoctoral researchers for the development of human resources in countries where these diseases are endemic.



Theileria(A)and Babesia(B) parasites | **Tick (A) and cattle (B) in Mongolia**



**Professor
Rika Umemiya-Shirafuji**

Ticks are obligatory hematophagous arthropods and are known as major vectors of various pathogens in vertebrates, such as *Babesia* and *Theileria* parasites. Our laboratory focuses on the molecular mechanisms underlying nutrient metabolism in unfed or fed ticks,

oogenesis, and pathogen transmission. We aim to contribute to the development of new methods for controlling ticks and tick-borne pathogens.

○ Nutrient metabolism in unfed ticks

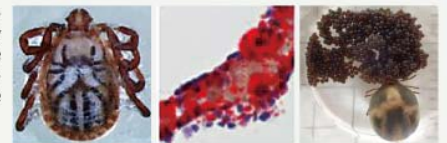
Most hard tick species have a life span of several months to years. Their life is essentially composed of relatively short parasitic periods and long non-feeding periods, without intake of blood. This remarkable viability is important for understanding the biology and physiology of ticks.

○ Nutrient metabolism in fed ticks

Energy and nutrient reserves provided by the digestion of a blood meal in female ticks allow the synthesis of vitellogenin (Vg), the yolk protein precursor. Synthesis and uptake of Vg are essential processes in the oogenesis of ticks.

○ Vector biology

Using ticks infected with parasites, under the laboratory conditions, we are studying the interactions between the transmission of parasites and the nutrient metabolism of ticks.



**Associate Professor
Thillaiampalam Sivakumar**

Bovine and equine piroplasmosis, caused by *Babesia* and *Theileria* species, are important tick-borne diseases affecting livestock health and productivity worldwide. Effective surveillance, accurate diagnosis,

and improved control strategies are essential. Our research focuses on the epidemiology, genetic diversity, and clinical impact of *Babesia*, *Theileria*, and related tick-borne pathogens in livestock. We conduct field studies to determine infection prevalence and identify risk factors, and perform molecular and phylogenetic analyses to better understand protozoal diversity and evolution. We also develop and improve diagnostic tools to support large-scale surveillance and work on *in vitro* cultivation to better understand protozoal biology and support improved control strategies. Through international collaboration, we contribute to enhanced disease surveillance, safer animal movement, and the development of effective control measures, while also supporting the training of young researchers in endemic regions.



First reported case of Babesia naookii infection in a cow | **Field survey of piroplasmosis** | **In vitro cultivation of equine piroplasma isolates from donkeys**



**Professor
Makoto Igarashi**

Toxoplasmosis is a zoonotic disease latently infecting 20% to 30% of the world's population. *Toxoplasma* causes serious problems in immune-suppressed people such as HIV patients and the elderly as well as in congenitally infected infants. This laboratory is working on unraveling the mechanism of *Toxoplasma's* parasitism in hosts.

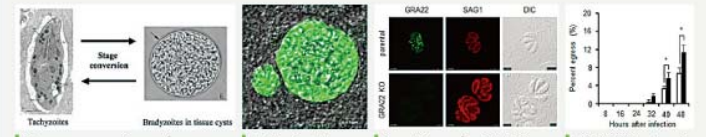
Main Research Project

○ Understanding *Toxoplasma's* parasitism in host cells:

We aim to identify target molecules for new medicines by isolating molecules involved in *Toxoplasma's* parasitism in host cells.

○ Understanding the transition process from acute to chronic *Toxoplasma* infection:

During the transition from an acute to a chronic infection *Toxoplasma* changes its life cycle from a propagation type to a cyst-forming type. By uncovering the mechanism of this change we aim to utilize it for vaccine development strategies.



Stage conversion of *Toxoplasma* | **Identification of the bradyzoite-specific DPA molecule and its expression in a cerebral cyst** | **Creation of a GRA22 gene knockout strain** | **Is GRA22 involved in egress from host cells?**



**Professor
Noboru Inoue**

As an WOAHP Reference Laboratory (WOAHP RL) Expert for Surra, I am in charge in periodic revisions of the WOAHP documents, such as Animal Health Code and the WOAHP Manual, in collaboration with other WOAHP RL Experts. I also provide definitive diagnosis of Surra at the request of international clients involved in the import and export of livestock, pets, and zoo exhibit animals, and conduct research on development of new diagnostic methods for trypanosomes.

We are conducting research to clarify the interaction between *Trypanosoma* and vectors/hosts at the molecular level. We particularly focused on the epimastigote (EMF) stage-specific cell surface molecules involving cell adhesion in tsetse salivary gland. We have so far discovered two EMF stage-specific surface proteins, CESP (function unknown) and TcEphBbR (hemoglobin receptor).

Development of simple/rapid diagnostic methods and social implementation research of epidemiological studies for practical use in endemic areas are being conducted. We have established Mongolian office and research facilities at the Institute of Veterinary Medicine, Mongolia, and are conducting collaborative research on dourine, which are serious concern in Mongolia, and are conducting joint research for the practical application of diagnostic methods and eradication measures for the protozoan diseases.



Our diagnostic kits are practically used in Mongolia | **WOAHP-NTTAT Network Meeting at WOAHP-HQ** | **Trypanosome sampling from dourine horse**



**Associate Professor
Keisuke Suganuma**

Trypanosomes are endemic in many countries and affect both humans and animals. However, no effective control measures are available for this disease. We therefore aim to develop and establish effective control strategies for trypanosomes.

Studies on developmental-stage conversion mechanisms. (Fig. 1)

Developmental-stage conversion (or cell differentiation) is essential for trypanosomes to parasitize mammalian hosts and vectors. Therefore, we aim to reveal the stage conversion mechanisms using gene manipulation technology, with the aim of developing new control strategies for trypanosomes.

Global surveillance and establishment of new laboratory strains of trypanosomes. (Fig. 2)

We aim to establish effective control strategies for trypanosomes in endemic countries based on field surveillance data. In addition, we intend to isolate and establish field strains of trypanosomes for further study.

Development of novel trypanocidal drugs.

We aim to find novel trypanocidal compounds by using *in vitro* drug screening systems and mouse models.

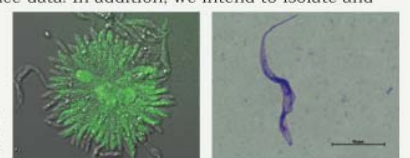


Fig. 1: GFP expressed transgenic *Trypanosoma congolense* (Suganuma et al., 2012 & 2013) | **Fig. 2: *Trypanosoma equiperdum* isolated from Mongolia (IVM-t1 strain) (Suganuma et al., 2016)**



**Professor
Shin-ichiro Kawazu**

Babesia

We are developing technology to manipulate genes expression using *Babesia*. So far, we have developed a foreign gene expression system (green fluorescent protein-expressing protozoa) and a gene knockout system, and we are currently trying to use live imaging to clarify the mechanism of the said protozoa's growth and infection into the red cells and the vector tick; and the pathology of cerebral babesiosis.

Schistosomiasis

Schistosomiasis is a zoonotic disease with major public and animal health implications in rural areas of developing countries. We are conducting comprehensive epidemiological surveys to eliminate the disease in the Philippines and Kenya. As a partner in the Southeast Asia-led Initiative (SEASED), our aim is to eliminate and monitor this infectious disease. To this end, we are conducting joint research into the development of ELISA and POCT.



Hyper (HzO2 sensor probe) expressing *Babesia bovis*

Schistosomiasis field survey in the Philippines.

Research Unit for Global Infection Control

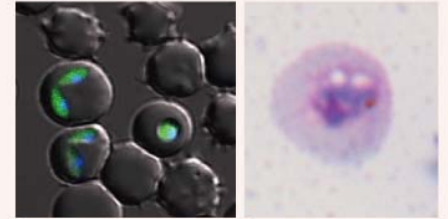
International Cooperation Unit



**Associate Professor
Masahito Asada**

Babesia and *Plasmodium* parasites are intraerythrocytic apicomplexan protozoans which cause severe morbidity and mortality to the animals and humans in the world. These parasites invade into the host erythrocytes and extensively modify structural and mechanical properties of the erythrocytes. To find a way to control babesiosis and malaria, we are studying on invasion and host erythrocyte modification mechanisms of the parasites by using genome manipulation techniques. Furthermore, we are performing field epidemiology of protozoan diseases.

- Mechanisms of parasite invasion into the erythrocyte
- Mechanisms of erythrocyte modification by the parasites
- Field epidemiology of protozoan diseases



Genome editing by CRISPR/Cas9 system | Ungulate malaria parasite



**Assistant Professor
Misuzu Okajima**

My research focuses on intraerythrocytic protozoan parasites that utilize rodents as hosts. I am currently conducting epidemiological surveys in Japan on *Babesia microti*, which is a zoonotic pathogen. Additionally, I am working to elucidate the mechanisms of chronic infection observed in multiple *Babesia* species using rodent *Babesia* species as primary models.

Furthermore, I have integrated the use of genetically modified mice into my research. By utilizing mice deficient in various physiologically active molecules, I aim to clarify at a molecular level which host defense mechanisms are critical for parasite elimination and how these parasites manage to evade the host's immune system.

[Main research projects]

- Epidemiological survey of rodent *Babesia* species in Hokkaido.
- Epidemiological survey of haemosporidia in wild birds across Japan.
- Analysis of protozoan pathogenic mechanisms using genetically modified mice.



Babesia microti

Establishing the infrastructure for the development of genetically modified mice

Research Unit for Innovative Medicine

Research Unit for Global Infection Control



**Professor
Yoshifumi Nishikawa**

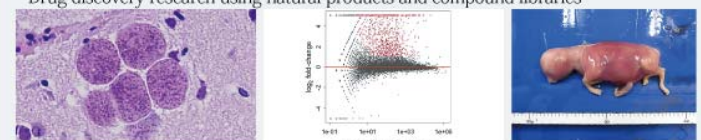
Lab HP



We are researching functional disorders of the central nervous system (CNS) and behavioral changes in host animals, as well as the mechanism of miscarriage or vertical transmission of protozoan infections. In addition, we are attempting to identify and analyze parasite-derived factors that control inflammatory response and immuno-suppression. Based on the results of this research, we are developing a new type of next-generation vaccine that can effectively transport a vaccine antigen to lymphoid tissues by utilizing lipid nanoparticles and can effectively stimulate immune cells. For practical application of our vaccine, we investigate the effects of the model vaccine based on infection models of mice and natural hosts.

[Main Research Projects]

- Subject of study: *Toxoplasma*, *Neospora*, *Sarcocystis*, *Cryptosporidium*, *Plasmodium*
- Study on behavioral changes of host animals and CNS disorder following parasite infection.
- Study on immune evasion mechanisms by parasite-derived proteins
- Study on the mechanism of reproductive disorder caused by parasite infection
- Development of new vaccine using lipid nanoparticles against protozoan disease
- Drug discovery research using natural products and compound libraries



Cyst of *Toxoplasma* in brain tissue

Comparative transcriptome using the brain tissue of a mouse infected with *Toxoplasma*

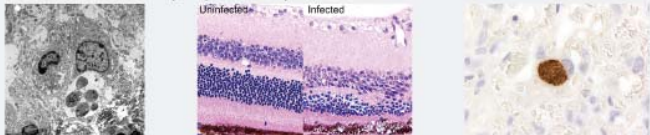
Aborted fetus from cow infected with *Neospora*



**Specially Appointed Assistant Professor
Nanako Ushio-Watanabe**

We are researching the pathology of parasite infections in animals and humans by using pathological analysis. Specifically, we focus on chronic infection in the central nervous system and acute and chronic infection in the retina caused by *Toxoplasma* and *Neospora*, as well as the infection during pregnancy.

- Chronic infection in the central nervous system: *Toxoplasma* and *Neospora* form cysts in the brain and cause lifelong chronic infections. Chronic infection with *Toxoplasma* has been reported to induce behavioral changes in the host. We are studying the pathogenesis and the impact of parasite-derived molecules that cause behavioral changes in the host.
- Acute and chronic infection in the retina: *Toxoplasma* infection, both congenital and acquired, can lead to inflammation in the retina, which result in blindness. We are focusing on Müller glial cells in the retina to analyze the pathogenesis of this condition.
- Infection during pregnancy: *Toxoplasma* and *Neospora* infections can cause miscarriage and neonatal infections. We analyze on mouse model of congenital infection and studying the pathogenesis of vertical transmission and postpartum depression.



Neospora caninum in a neuron.

Degeneration of the retina

Toxoplasma gondii in a trophoblast.

Research Unit for Innovative Medicine

Research Unit for Innovative Medicine

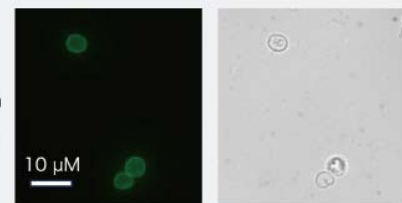


**Specially Appointed Assistant Professor
Rie Kubota**

We are researching the cellular parasitic mechanisms of *Cryptosporidium*, which causes diarrhoea in humans and animals. *Cryptosporidium* parasites infect all mammals including humans and animals. There is no effective vaccine or prophylactic drug, and *Cryptosporidium* infection, especially in calves, causes lethal diarrhoea, resulting in economic losses to the cattle industry, including Japan. Parasites invade intestinal epithelial cells and shed their seeding (oocysts) by sexual reproduction, but the detailed mechanisms of parasitism and development are not clear. We aim to understand the parasitic strategies of *Cryptosporidium* parasites, including the analysis of their gene function, using genetic modification techniques.

[Main Research Projects]

- Elucidation of the cellular invasion mechanism of *Cryptosporidium* parasite
- Discovery of anti-cryptosporidium parasite drugs
- Development of in vitro culture systems for *Cryptosporidium* parasite
- Study on the intestinal microflora associated with *Cryptosporidium* infection
- Discovery of vaccine antigens



Cryptosporidium oocysts from bovine faeces (immunofluorescence staining)

10 μM

Base of Activities

As an international joint research center for the control of protozoan diseases, the National Research Center for Protozoan Diseases develops technology for the diagnosis, prevention and treatment of livestock protozoan diseases in domestic and foreign countries. We utilize an experimental system that simulates the life cycle of each protozoon in nature (experimental system of authentic infection). In addition, we use WOAH Collaborating Centre-related research to promote developmental research into technology relating to protozoan disease prevention that will

become the international standard. In the 4th mid-term target period, we are conducting a national project entitled "Establishment of a comprehensive drug discovery research center utilizing the analysis matrix for livestock protozoan diseases (Drug Discovery Project)". For the development of new therapeutic agents for livestock protozoan diseases, screening of candidate compounds and studies of their mechanism of action are underway. The goal of this project is establishment of "research center for drug discovery on livestock protozoan diseases" with a view to realizing social implementation.



Online International Symposium on Ticks as a Joint Usage/Research Center Project



Joint research laboratory established in Makerere University, Uganda



Joint laboratory established in Shanghai Veterinary Research Institute, China



Joint research laboratory established in the Institute of Veterinary Medicine in Mongolia

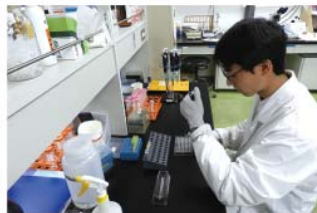
Social Contribution Activity

We offer diagnostic services for infectious diseases, especially protozoan diseases, in domestic animals in cooperation with the Veterinary Medical Center at Obihiro University of Agriculture and Veterinary Medicine to help the community with our research achievements. We also provide extremely advanced and specialized diagnostic services for protozoan diseases that have been developed by the National Research Center for Protozoan Diseases. Moreover, as a core research institute for protozoan

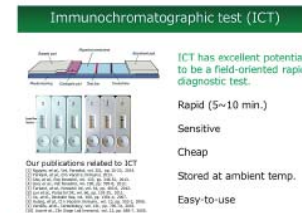
diseases, we would like to contribute to the development of protozoan disease research worldwide. We therefore provide bioresources such as protozoan parasites, ticks, cDNA libraries, and monoclonal antibodies. Information about the bioresources offered is openly available on our home page. Furthermore, by establishing and providing systems of bioassay for pathogenic protozoan parasites, we contribute actively to development of anti-protozoal agents.



Introduction of our research to the general public



Special diagnostic service for infectious diseases



Immunochromatography for the diagnosis of protozoan diseases



Bioresources (protozoan parasites)

Situation for Acquisition of External Funds (number)

Source of Budget	Budget Item	FY2023	FY2024	FY2025
Japan Society for the Promotion of Science	Grant-in-Aid for transformative research areas (A)	0	1	1
	Grant-in-Aid for Scientific Research (B)	6	6	5
	Grant-in-Aid for Scientific Research (C)	1	1	1
	Challenging Research (Exploratory)	2	2	2
	Grant-in-Aid for Young Scientists	1	1	2
	Fostering Joint International Research (B)	4	3	3
	International Collaborative Research	1	2	1
	Grant-in-Aid for JSPS Fellows	6	3	0
	Core-to-Core Program	1	1	1
	Joint Research Projects/Seminars	2	1	1
Ministry of Agriculture, Forestry and Fisheries	Bilateral joint research project	1	1	0
Japan Agency for Medical Research and Development	Research Program on Emerging and Reemerging Infectious Diseases	1	1	1
	Japan Program for Infectious Diseases Research and Infrastructure	1	1	1
JICA - JST SATREPS	Science and Technology Research Partnership for Sustainable Development	0	1	1
Private Research grant etc. (500,000 yen or more)	Joint research	1	1	2
	Contract research	3	3	0
	Grants	2	5	4
Total		32	33	25

The Journal of Protozoology Research

Introduction of Publications

This journal has been issued once or twice a year as an international journal on protozoan diseases since 1990.

Call for papers

We are calling for review papers, original papers and short communication papers pertaining to the study of protozoan diseases, protozoology, and arthropod vectors.

Contact regarding contribution to this journal

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Members of the Center

◆ **Director General** Noboru Inoue **Vice Director General** Yoshifumi Nishikawa

◆ **Department of Drug Discovery and Development**

Research Unit for Innovative Medicine

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Specially Appointed Assistant Prof. Nanako Ushio-Watanabe

Specially Appointed Assistant Prof. Rie Kubota

◆ **Department of Disease Control**

Research Unit for Molecular Diagnostics

Prof. Naoaki Yokoyama / Prof. Rika Umemiya-Shirafuji

Assoc. Prof. Thillaiampalam Sivakumar

Research Unit for Advanced Preventive Medicine

Prof. Noboru Inoue / Assoc. Prof. Keisuke Sukanuma

Research Unit for Infection and Pathology

Prof. Makoto Igarashi

◆ **Department of Global Cooperation**

Research Unit for Global Infection Control

Assoc. Prof. Masahito Asada / Assistant Prof. Misuzu Okajima

Research Unit for Global Surveillance

Prof. Naoaki Yokoyama (Concurrent)

Assoc. Prof. Keisuke Sukanuma (Concurrent)

International Cooperation Unit

Prof. Shin-ichiro Kawazu / Prof. Makoto Igarashi (Concurrent)

Prof. Noboru Inoue (Concurrent)

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Honorary Prof. DeMar Taylor

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Technical Assistant Tsuyoshi Habaguchi, Manami Yanagihara

Assistant Clerk Yuko Kanomata, Nozomi Suzuki, Yumiko Yamamoto

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Shigeyuki Kano

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Yasuhiko Suzuki

Nariaki Nonaka

Badgar Battsetseg

Kenji Hirayama

Toshihiro Horii

Hong Yeonchul

Makoto Igarashi

Noboru Inoue

Shin-ichiro Kawazu

Yoshifumi Nishikawa

Naoaki Yokoyama

Rika Umemiya-Shirafuji

Member of the collaborative researchers in 2026

(in order of the Japanese syllabary, with titles omitted)

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Hiroimi Ikadai

Yuzuru Ikehara

Takahiro Ishizaki

Akihiro Ochi

Hiroki Kaneko

Satoru Kawai

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Tatsuki Sugi

Takeshi Suzuki

Masakatsu Taira

Kiyotada Naito

Youichi Nakao

Coh-ichi Nihei

Kenji Hikosaka

Akikazu Fujita

Tatsunori Masatani

Keita Matsuno

Shingo Miyawaki

Toshihiro Murata

Ayumu Moriya

Junya Yamagishi

Jun Yamamoto

Ryuji Yanase

Yasunaga Yoshikawa

Apinya Arnuphappasert

Atambekova Zhylidy

Atefeh Fathi

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Bernard Cheruiyot Rono

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Liqing Ma

Maria Cecilia Venturini

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Sanjay Kumar

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Japan Institute for Health Security (JIHS)

Professor, The University of Tokyo

The Institute of Medical Science

Distinguished Professor, Hokkaido University

International Institute for Zoonosis Control

Professor, Hokkaido University School /

Faculty of Veterinary Medicine

Director, Institute of Veterinary Medicine

Professor, Nagasaki University School of

Tropical Medicine and Global Health

Professor, Osaka University

Research Institute for Microbial Diseases

Professor, Kyungpook National University

Professor, NRCPD

Professor, NRCPD

Professor, NRCPD

Professor, NRCPD

Professor, NRCPD

Professor, NRCPD

Chief Researcher, Japan Institute for Health Security

Professor, Kitasato University

Professor, Chiba University

Senior Assistant Professor, Rakuno Gakuen University

Researcher, Japan Racing Association

Professor, Hamamatsu University School of Medicine

Professor, Dokkyo Medical University

Assistant Professor, Chiba University

Researcher, Japan Institute for Health Security

Assistant Professor, Hokkaido University

Professor, Tokyo University of

Agriculture and Technology

Chief Researcher, Japan Institute for Health Security

Associate Professor, Kagoshima University

Professor, Waseda University

Senior Researcher, Microbial Chemistry Research Foundation

Associate Professor, Chiba University

Professor, Kagoshima University

Associate Professor, Gifu University

Associate Professor, Hokkaido University

Associate Professor, Gifu University

Associate Professor, Tohoku Medical and

Pharmaceutical University

Senior Researcher, Hokudo Co., Ltd.

Professor, Hokkaido University

Assistant Professor, The University of Texas

Southwestern Medical Center

Chief Researcher, Japan Institute for Health Security

Associate Professor, Kitasato University

Senior Assistant Professor, Rajamangala

University of Technology Srivijaya

Researcher, Kyrgyz Research Institute of Veterinary

Named After A. Duisheev,

Post doctoral, The University of Glasgow

Director, Institute of Veterinary Medicine

Researcher, Kyrgyz Research Institute of Veterinary

Named After A. Duisheev

Senior Principal Veterinary Officer, Kenya Wildlife Service

Professor and Chair in Parasitology, University of Otago

Research Scientist, Institute of Parasitology,

Biology Centre CAS

Assistant Professor, Kansas State University

Veterinary Research Officer, Veterinary Research Institute

Professor, Shanghai Veterinary Research Institute,

Chinese Academy of Agricultural Sciences

Postdoctoral researcher, University of Antwerp

Associate Professor, Tribhuvan University

Professor, Qinghai University Qinghai Academy of

Animal Sciences and Veterinary Medicine

Head Professor / LAINPA Director-Researcher,

La Plata National University, Faculty of Veterinary Sciences

Senior professional scientist, Mahidol University

Faculty of Tropical Medicine,

Principal Scientist, ICAR-National Research Centre

on Equines, India



HISTORY

I The Laboratory for Protozoan Immunology (1984-1990)

April 1984. The Laboratory for Protozoan Immunology was established as an annex of the Department of Veterinary Physiology (Prof. Em. Naoyoshi SUZUKI)

II The Research Center for Protozoan Molecular Immunology (1990-2000)

June 1990 The Research Center for Protozoan Molecular Immunology granted permission by MEXT, and established as a Joint Research-Educational Facility at Obihiro University until March 31, 2000. Research Unit of Molecular Immunology was established.

April 1992 Research Unit of Pathophysiology, established.

June 1993 New research building constructed with 462m² area.

April 1995 Research Unit of Disease Control and Genetics, established.

April 1997 Research Unit of Molecular Arthropodology, established.

November 1997 New research building constructed with 970m² area.

III National Research Center for Protozoan Diseases (2000 to the present)

April 2000 Establishment of the "National Research Center for Protozoan Diseases:NRCPD" as a national research facility covering a 10 year period (2000-2010). Research Unit for Molecular Diagnosis and Research Unit for Advanced Preventive Medicine, established.

March 2002 Extension of research building completed with 1,730 m² area.

October 2002 NRCPD recognized and selected to implement the "21st Century Center of Excellence (COE) Program", by MEXT.

April 2003 Department of Large Animal Infections, established.

April 2005 Department of Global Surveillance of Protozoan Diseases, established with the three Research Units.

March 2006 Extension of research building completed with 1,520 m² area

June 2007 Certified as an WOAHP Reference Laboratory (bovine babesiosis and equine piroplasmosis: Prof. Igarashi, surra: Prof. Inoue).

May 2008 Certified as an WOAHP Collaborating Centre (the first facility in the world in the field of protozoan diseases).

June 2009 Certified as the Joint Usage Research Center by MEXT.

November 2012 Bio-Self Regulating Science Laboratory (Hakuju donated fund laboratory), established.

March 2013 Research Unit for Global Infection Control, established (Tenure-Track Promotion Program, JST).

April 2016 Re-certified as the Joint Usage Research Center by MEXT.

March 2017 Certified as ISO/IEC17025:2005

(Bovine babesiosis, Equine piroplasmosis, and Surra).

January 2018 Research Unit for International Surveillance, established

April 2022 Re-certified as the Joint Usage Research Center by MEXT.

April 2022 Department of Drug Discovery and Development,

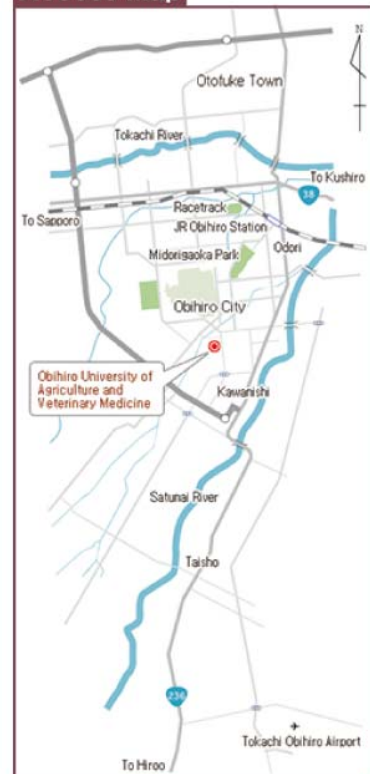
Research Unit for Innovative Medicine, established

June 2025 Re-designated as a WOAHP Collaborating Centre

Access to us



Access Map



From Obihiro Station to the University

Tokachi Bus

There are 2 bus routes from Obihiro Station to the University.

① From Obihiro Station Bus Terminal #9, take bus #70 or #72 bound for Oozora Danchi.

Get off at **Ryokuyou Koko Mae Bus Stop**. (about 30 min.)

It's about a 15 min. walk to the University entrance.

② From Obihiro Station Bus Terminal #9, take bus #73 bound for Suzuran Plaza.

Get off at **Nougyou Koko seimon kita Mae Bus Stop**. (about 30 min.)

It's about a 15 min. walk to the University entrance.

Taxi

It takes about 20 min. and costs around 2,400 yen (about 7 km).

From Tokachi-Obihiro Airport to the University

Airport shuttle Bus

There is a bus from the airport to Obihiro Station. (about 40 min., 1,000 yen)

From Obihiro Station see the information shown above.

Taxi

It takes about 30 min. by taxi and costs around 7,000 yen (about 21 km).



▲Tokachi Bus



▲Airport shuttle bus

