

# NRCPD-OUAVM Joint Research Report

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## 1. Principal investigator

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Position: Lecturer

Affiliation: Rajamangala University of Technology Srivijaya

## 2. Project title:

Isolation of *Babesia bovis* of Thai origin and characterization of the VESA gene family encoding its virulent factors

## 3. Collaborating research group members at NRCPD

Name: ASADA Masahito

Position: Associate Professor

## 4. Research period (in mm/dd/yyyy, and total number of years)

April 1, 2024 – March 31, 2025

## 5. Purposes and objectives

This study aims to establish cryostabilates of *Babesia* parasites and genetically characterize genes related to virulent factors, specifically the VESA gene family, from Asian isolates of *Babesia bovis*, providing valuable insights into parasite biology. The parasite cryostabilates acquired in this study will be valuable for future research, including genome-wide studies, as well as the development of drugs.

## 6. Outline of research process

### Research in Thailand

#### 1. Blood sample collection:

Blood samples were collected from 15 cattle that exhibited *Babesia*-like clinical signs, including depression, loss of appetite, anemia, dark-colored urine, and a history of contact with ticks, the vector of *Babesia bovis*. The blood was added cryopreserve reagent and stored in liquid nitrogen.

#### 2. Blood smear screening:

Blood samples were smeared onto slides, fixed with methanol, and stained with 10% Giemsa solution. The slides were then examined under a light microscope (1000x magnification) to identify the piriform



shape, which is characteristic of Babesia.

### 3. DNA extraction:

DNA was extracted from 200 µL of each blood sample using a commercial kit (NucleoSpin Blood Extraction Kit).

### 4. PCR screening:

Fifteen samples were screened by PCR using the primers BTH 18S 1st F (GTGAAACTGCGAATGGCTCATTAC) and BTH 18S 1st R (AAGTGATAAGGTTACAAAATTCC) for the first round of PCR. The second round of PCR used primers BTH 18S 2nd F (GGCTCATTACAACAGTTATAGTTTATTTG) and BTH 18S 2nd R (CGGTCCGAATAATTCACCGGAT), following the protocol described by Masatani et al. (2017).

Only PCR-positive samples had their first-round PCR products prepared and sent to the National Research Centre for Protozoan Diseases (NRCPD) at Obihiro University of Agriculture and Veterinary Medicine, Japan, for further analysis.

### Research at NRCPD

#### 1. DNA Sequencing:

PCR products were re-amplified using the second-round primers and a larger reaction volume (50 µL). The predicted PCR bands were excised and purified using a gel purification kit (NucleoSpin PCR Clean-up and Gel Purification Kit). The purified products were then subjected to sequencing.

#### 2. Nucleotide Analysis:

The obtained sequences were edited using BioEdit software and compared against known sequences using the nBLAST tool.

### 7. Outline of research achievements

Out of 15 samples, 6 samples were positive from PCR (about 1,500 bp). The result from sequencing revealed that three samples were *B. bovis*, two samples were *B. bigemina* and one sample was *T. orientalis*. There were two tubes of *B. bovis* of Thai strain were stored in liquid nitrogen.

### 8. Publication of research achievements

#### Reference

Masatani, Tatsunori, et al. "Detection and molecular characterization of Babesia, Theileria, and Hepatozoon species in hard ticks collected from Kagoshima, the southern region in Japan." Ticks and tick-borne diseases 8.4 (2017): 581-587.